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STRUCTURE OF CYTOSININE, A STRUCTURAL COMPONENT OF BLASTICIDIN S

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In previous papers^{1,2}, we have reported the isolation and characterization of a new nucleoside designated cytosinine from the hydrolysis products of blasticidin S, a useful antibiotic against rice blast desease. We wish to present the details leading to the assignment of structure I to cytosinine.

Cytosinine (I), $C_{10}H_{12}O_4N_4$, m.p. 244-245°, $[\alpha]_{D}^{18}$ -20°(c, 1 in H₂O), pKa 2.4, 4.6 and 8.0, λ_{max}^{01NHC} 274 mµ (£ 13,800), $\lambda_{max}^{01N,NaOH}$ 267 mµ (£ 7,500), showed the existence of two ethylenic groups (NME (in D₂O) ppm: 7.72 and 6.14 (2H d, AB type, J = 7.5 cps) and 6.45-6.00 (2H broad)), two amino groups (Van Slyke determination) and one carboxylic group. By the action of methanol containing 3% hydrogen chloride, I gave corresponding methyl ester dihydrochloride, $C_{10}H_9O_3N_4(OCH_3)$ ·2HCl,

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m.p. 220-223°(dec.), IR (Nujol) 1730 and 1230 cm⁻¹, pKa 4.6 and 8.0, which was acetylated with acetic anhydride and triethylamine to afford N, N'-diacethyl methylester (III), $C_{10}H_{9}O_{3}N_{4}$ (OCH₃)·(CH₃CO)₂, m.p. 263-266°.

UV spectral evidence indicated probable presence of cytosine nucleus, on which a substituent attached at N-1 position.

Acid hydrolysis of I gave one mole each of cytosine, levulinic acid, ammonia and carbon dioxide as characterizable products, whereas further attempt to cleave the glycosidic linkage of I was unsuccessful without extensive decomposition.

In an effort to gain information regarding the sugar portion^{*1}, reduction and oxidative cleavage of ethylenic function in I were undertaken. On reduction with PtO₂ in acetic acid, I gave a mixture of the dihydrocytosinine (IV), $C_{10}H_{14}O_4N_4$, m.p. 227-229°(dec.), NMR (D₂O) ppm: 7.25 and 5.45 (2H d, J = 7.5 cps), 5.10 (1H m), 3.50 (1H d, J = 10.5 cps), 2.7C (1H m) and 1.40 (6H m), hydrogenolysis product (V), $C_6H_{11}O_3N$, m.p. 275-278°, and cytosine. The facile hydrogenolysis of I to cytosine and V indicated that an ethylenic function in C₆ molety located at allylic position to the glycosidic C-N bond.

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The presence of sugar moiety was presumed, since I gave levulinic acid on acid hydrolysis.

V characterized as monobasic monoacidic compound of pKa 9.2 and 2.4 gave corresponding N-acethyl methylester, $C_{6}H_{9}O_{2}N$. (OCH₃).(CH₃CO), m.p. 130°, IR (Nujol) 1740, 1670 and 1230 cm⁻¹. The IR spectra of V and deuterated V exhibiting characteristic bands of tetrahydropyran³ at 1100 and 1060 cm⁻¹ (C-O-C stretching vibration) established the presence of tetrahydropyran ring.

Ozonolysis of III in acetic acid followed by oxidation with hydrogen peroxide and acid hydrolysis furnished β -hydroxy aspartic acid (VI), $C_4H_7O_5N$, m.p. 287-288°(dec.), $[\alpha]_p^{20}$ -47° (c, l in H₂O). On diazotization, VI afforded meso-tartaric acid, therefore, the erythro-D-configuration⁴ was established. VI was a substantially important structural fragment of the sugar portion in I and was also obtained from blasticidin S itself by a similar ozonolysis.



These results established the structure of V as 3-aminotetrahydropyran-2-carboxylic acid. On the basis of these accumulated evidence the structure of cytosinine was elucidated as formula I.

Strepeochemical feature of I was next to be considered, e.g. the configuration of erythro-D- β -hydroxy aspartic acid provided evidence for the (R)-configuration at C₄ and C₅' respectively. The NMR spectrum of IV showing a distinct doublet at 3.50 ppm (lH, J = 10.5 cps) assigned for C₅' proton established the C₄'-C₅' protons to be diaxial. The configuration of glycosidic linkage remained uncertain, but it was presumed to be β form, since nucleosides from natural source were reported as β .

It was necessary to consider the mechanism of levulinic acid formation from I. Transformation of hexoses into levulinic acid by the action of acids was well established⁵. Similarily, the formation of levulinic acid, ammonia and carbon dioxide from I could be explained as follows:



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References

- S. Takeuchi, K. Hirayama, K. Ueda, H. Sakai and H. Yonehara, <u>J. Antibiotics</u>, <u>11A</u> 1 (1958).
- H. Yonehara, S. Takeuchi, N. Otake, T. Endo and Y. Sumiki, J. Antibiotics, <u>16A</u> 195 (1963).
- 3. S. C. Barket and R. M. Badger, <u>J. Chem. Soc</u>., 4397 (1950).
- 4. H. J. Salbach, <u>J. Biol. Chem.</u>, <u>229</u> 437 (1957).
- 5. H. P. Teunissen, <u>Rec. Trav. Chim.</u>, 50 1 (1930).